Extraction Site Preservation Using New Graft Material That Combines Mineralized and Demineralized Allograft Bone: A Case Series Report with Histology

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Abstract: This retrospective case series reports on the use of a new allograft bone product composed of a 70:30 ratio of mineralized to demineralized cortical bone particles to preserve the alveolar ridge dimensions of patients requiring tooth extraction with plans for future dental implant placement. Ten patients received atraumatic tooth extractions with subsequent placement of the blended bone allograft. All sites were covered with a single layer of amnion-chorion, which was intentionally left exposed. After an average healing time of 14 weeks, the surgical sites were accessed for placement of dental implants. Prior to implant placement, trephine core samples were taken for histologic analysis. Dental implants were successfully placed in all previously grafted surgical sites; no sites required any additional bone augmentation. Histologic analysis of trephined bone core samples revealed bone compositions similar to those described in previously published site preservation studies. The results of this case series suggest that blended bone allograft containing a 70:30 ratio of mineralized to demineralized cortical bone particles can be successfully used to facilitate future placement of dental implants with as little as 14 weeks of healing.

LEARNING OBJECTIVES

- discuss the physiologic changes in osseous structure that result from tooth extraction and how they impact the potential placement of dental implants

- explain the rationale behind extraction site preservation with bone allograft if implant placement is planned

- describe the basis for the use of a blended bone allograft product containing both mineralized and demineralized cortical bone for extraction site preservation

Replacement of missing teeth with dental implants now ranks among the most successful procedures in dentistry,[1] with millions of implants being placed annually. Achieving optimal esthetics, function, and phonetics with implant-supported restorations depends largely on the presence of adequate bone at the dental implant placement site.

It is well documented that following tooth extraction, reductions in both the alveolar ridge height and width typically occur. The processes of tissue modeling and remodeling have been investigated in both animals[2] and humans,[3,4] and the routine loss of ridge volume has been explained by the absence of daily stress/strain stimulus required for physiological maintenance of the bone anatomy.[5] In the absence of site preservation following tooth extraction, significant tissue-contour loss occurs during the 3 months post-extraction, averaging 3 mm to 5 mm at 6 months.[6,7] Horizontal bone resorption of up to 30% lingually and 56% buccally has been reported,[8] with overall reduction in ridge width of up to 50% found 1 year after extraction.[3]

To reduce the dimensional ridge changes that occur during post-extraction healing, various site-preservation techniques have been proposed,[9-13] and significant reductions of the dimensional changes have been reported.[14,15] Detailed analysis of the 2003 Iasella study,[14] for example, demonstrates that approximately 125% more loss of horizontal ridge width can be expected in the absence of site preservation (1.2-mm ridge loss with site preservation versus 2.6-mm ridge loss without site preservation), while up to 244% more loss of vertical ridge height can be expected with no site preservation
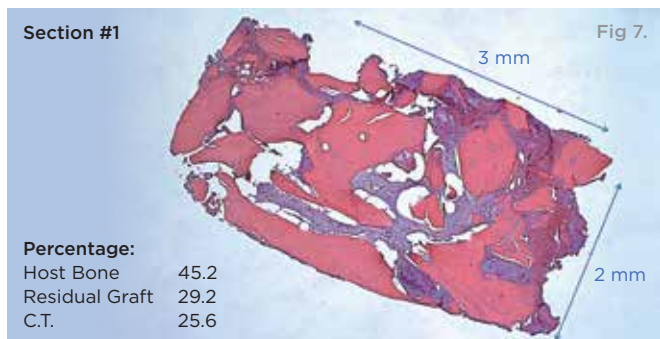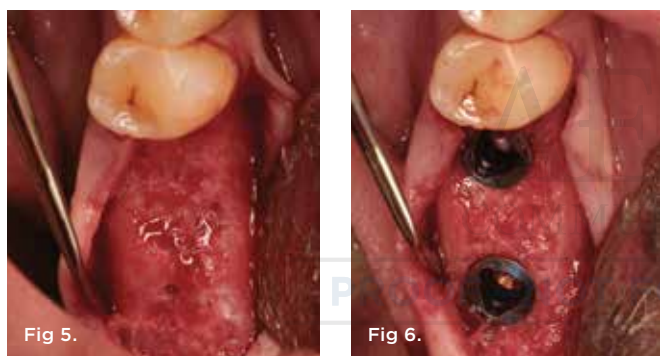
Fig 1.

Fig 2.

Fig 3.

Fig 4.

Fig 5.

Fig 6.

Section #1

Fig 7.

3 mm

2 mm

Percentage:
Host Bone   45.2
Residual Graft   29.2
C.T.   25.6

Fig 1. Case 1: Hopeless mandibular first and second molars prior to extraction. **Fig 2.** Atraumatic extraction of hopeless mandibular first and second molars. **Fig 3.** Placement of bone allograft into extraction sites. **Fig 4.** Intentional non-primary closure of surgical sites. **Fig 5.** Healed bony ridge at 14 weeks following site preservation. **Fig 6.** Placement of dental implants into healed bone. Note complete circumferential bone surrounding all aspects of the dental implants. **Fig 7.** Low-power hematoxylin and eosin stain of trephine bone core sample from Case 1 showing conglomeration of vital bone, residual bone allograft, and connective tissue.

(1.3-mm gain in ridge height with site preservation versus 0.9-mm loss of ridge height without site preservation). Previously published approaches for extraction site preservation include placement of allograft, xenograft, and alloplast graft materials, with or without the use of occlusive membranes. For allografts, both mineralized and demineralized bone have been utilized. However, no mixtures of mineralized and demineralized allograft have generally been commercially available until recently. One possible explanation for this is that US Food and Drug Administration (FDA) regulations require that all human bone allograft products be labeled with unique alphanumeric identification codes that allow each manufacturer to record and track the graft material to its recipient (and vice versa).[16] Consequently, offering a product that includes bone from the same donor that has been processed in two different ways requires a somewhat more complicated manufacturing process.

The aim of this case series is to document the use of a relatively new bone allograft product comprised of blended mineralized and demineralized cortical bone for extraction site preservation procedures.

Case Series Treatment

Ten consecutive patients who had sought treatment at a private practice limited to periodontics and dental implants in Austin, Texas, were identified. All patients had required extraction of a tooth with site preservation to facilitate future placement of dental implants. Written and signed informed consent was obtained prior to treatment for all patients. Local anesthesia was obtained using 4% articaine hydrochloride with 1:100,000 epinephrine (Septodont, www.septodontusa.com). In all cases, a full-thickness mucoperiosteal flap was reflected, and tooth removal was accomplished atraumatically. Multi-rooted teeth were sectioned with a high-speed handpiece, while single-rooted teeth were removed with periotome assistance.

Upon removal of the tooth, sockets were thoroughly degranulated with hand instruments and irrigated with sterile saline. Each socket was then filled with CREOS 70/30 bone allograft (Nobel Biocare, www.nobelbiocare.com) and covered with a single layer of amnionchorion (Snoasis Medical Products, www.snoasismedical.com). No primary closure was attempted in any cases of this report. In cases with significant osseous defects such as a missing buccal plate, a secondary collagen membrane (Community Tissue Services™, www.communitytissue.org) was utilized for graft containment at the buccal wall, and an amnion-chorion barrier was perpendicularly layered over the collagen membrane. The purpose of the amnionchorion membrane in all cases, whether used alone or in conjunction with a collagen barrier, was to act as a matrix for rapid epithelial cell migration over the non-primary closure of the socket. Prior to placement, the bone allograft was only hydrated with sterile saline, and no growth factors were used or mixed with the bone. To reiterate, in all cases, primary closure was intentionally not achieved.

Post-surgically, all patients were prescribed antibiotics depending on their various individual drug allergies and a combination of analgesic and narcotic medications for pain. Oral rinses such as chlorhexidine were not prescribed following surgery, and patients were instructed to brush and floss normally. Patients returned for an initial follow-up visit 10 days post surgery, at which time sutures were removed and teeth were cleaned with hand instruments. Six

weeks following surgery, patients were seen for an interim visit that included radiographic and clinical examination of the surgical site.

Twelve to 16 weeks after the initial surgery (average 14.2 weeks), each patient presented for dental implant placement. While preparing the previously grafted sites for implant insertion, a 2-mm trephine core sample was taken from each surgical site prior to completion of the final drilling sequence. The trephined bone cores were submitted to the University of Missouri–Kansas City School of Dentistry for histologic examination.

Results

This article reports on treatment of 10 patients with a new blended bone allograft material that was placed in fresh extraction sockets to preserve the alveolar ridge in anticipation of subsequent implant placement. A total of six females and four males with an average age of 54.6 years (± 6.2 years) were included in this case series. Two of the patients were smokers, and one patient was a well-controlled diabetic. All other patients had relatively unremarkable medical and social histories. The 10 surgical sites consisted of three maxillary molars, three mandibular molars, two maxillary premolars, one mandibular premolar, and one maxillary central incisor. All 10 surgical sites healed uneventfully with no incidences of infection, graft loss, or other complications.

After roughly 3 months (14.2 weeks), bone formation in all cases was adequate in both buccolingual and apicocoronal dimensions to enable proper locational placement of the planned dental implants without the need for any additional bone-graft material. Figure 1 through Figure 19 depict three of the cases. Analysis of the trephine core samples (Table 1) revealed results consistent with findings of previously published site-preservation studies.

Discussion

Bone allografts have been used in dentistry for over 40 years, with more than 800,000 transplantations performed annually in the United States, a more than 400% increase since 1972.[17] Rigorous donor screening and aseptic proprietary processing programs have rendered their use safe and effective.[16] All harvested tissue is routinely tested for human immunodeficiency virus (HIV), hepatitis B and C, human T-lymphotropic virus, as well as bacterial and fungal contaminants. Once harvested, the bone is processed in hyper-clean facilities that strictly control temperature, humidity, ionization, electrostatic discharge, air pressure, air ventilation, and air filtration.[16] Soft tissue is stripped away, and the hard tissue is then sectioned to manageable sizes, rigorously cleansed, and decontaminated. Nearly all of the moisture content is eliminated via lyophilization or repetitive solvent baths that reduce antigenicity and enable long shelf storage of up to 3 years at room temperature.

The particle size of mineralized bone allograft typically ranges between 250 to 1,000 micrometers. Once reduced and packaged, additional low-dose irradiation may also be administered for terminal sterilization of the graft. In the case of demineralized bone allograft, the bone is typically immersed in a hydrochloric acid bath for various lengths of time in an effort to demineralize the bone by reducing calcium content. The bone is then washed in various proprietary buffer solutions to remove residual acid prior to terminal processing with
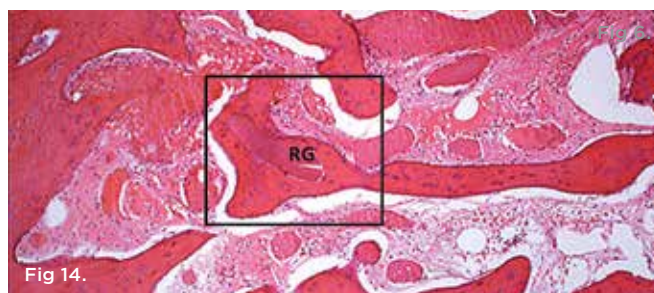

Fig 8.

Fig 9.

Fig 10.

Fig 11.

Fig 12.

Fig 13.

Fig 14.

Fig 8. Case 2: Hopeless mandibular first molar prior to extraction. **Fig 9.** Note the significant osseous defect in the buccal osseous wall following atraumatic tooth extraction. **Fig 10.** Placement of bone allograft into extraction site with collagen membrane on buccal aspect for graft containment. **Fig 11.** Intentional non-primary closure following the placement of an amnion-chorion barrier over the bone allograft. **Fig 12.** Note the significant formation of bone and repair of the buccal osseous defect at 3 months healing. **Fig 13.** Placement of dental implant into healed bone. Note complete circumferential bone surrounding all aspects of the dental implant. **Fig 14.** Low-power hematoxylin and eosin stain of trephine bone core sample from Case 2 showing residual bone allograft particle (RG) surrounded by vital bone.

low-dose gamma irradiation. These tissue processes vary depending on the proprietary formulation of individual bone banks. The bone allograft used in this particular case series was processed via the patented Allowash® (LifeNetHealth, www.accesslifenethealth.org) process and had a particle size of 250 to 1,000 micrometers. Additionally, it was a blended ratio of 70:30 mineralized to demineralized cortical bone from the same individual donors.

Multiple studies have evaluated the bone-forming effectiveness of both mineralized and demineralized bone allografts when used for dental surgery.[18-24] Different mechanisms are at work in the two types of graft materials.[25] Demineralized bone allograft, for example, provides a source of osteoinductive factors.[26] It contains type I collagen and various proteins such as bone morphogenetic protein (BMP) with the capacity to influence cell behavior, including proliferation and attachment.[27] Shigeyama and colleagues tested commercially prepared demineralized bone allograft via multiple assays and Western Blot analysis, with results showing the allograft retained proteins such as BMPs 2, 4, and 7.[27] By reducing the calcium content of allograft particles via degranulation, it is thought that faster release of residual proteins such as BMPs may be attained for improved bone healing. Multiple studies have documented higher osteogenic capacity in vitro for demineralized bone

allograft compared to mineralized bone allograft.[18,19] Accordingly, one recent study that compared demineralized to mineralized bone allograft used in extraction socket site preservation found significantly greater new bone formation from the use of the demineralized material.[28] In this study, however, it is interesting to note that no significant differences in alveolar ridge dimensional changes were noted between the two allograft groups and that only sockets with minimal dehiscence defects were included in the study. The latter statement highlights one of the problems that many clinicians find with demineralized bone allograft. Because the product is demineralized, its structural integrity is minimized. While this is beneficial for the rapid release of BMPs, it may pose a problem in osseous defects that are not well contained, such as extraction sockets with significant dehiscences or absent bony walls. The issues associated with allograft structural integrity are minimized with mineralized products. Because they retain their calcium content, mineralized cortical allografts are considered osteoconductive materials[29] and are better suited for defects with less than optimal containment, especially when compared with demineralized allografts.[30]

The availability of a product that combines both demineralized and mineralized bone allograft allows the benefits of both to be obtained. Because the cases in this series were consecutive, there

Fig 15. **Fig 15.** Case 3: Hopeless mandibular molar prior to extraction. **Fig 16.** Note the significant dehiscence defect in the buccal wall prior to tooth extraction. **Fig 17.** Placement of bone allograft into the extraction site. **Fig 18.** Surgical site at 3 months healing. **Fig 19.** Placement of dental implant into healed bone. Note complete circumferential bone surrounding all aspects of the dental implant.

TABLE 1

Histologic Analysis of Trephined Core Bone Samples Following Healing from Extraction Site Preservation

SITE	HEALING TIME (WEEKS)	VITAL BONE (%)	GRAFT MATERIAL (%)	CONNECTIVE TISSUE (%)
Mandibular molar	12	57.9	15.5	26.6
Mandibular molar	14	45.2	29.2	25.6
Mandibular molar	14	51.1	24.2	24.7
Maxillary molar	14	35.2	29.2	35.6
Maxillary molar	15	39.8	34.3	25.9
Maxillary molar	15	36.2	38.3	25.5
Maxillary premolar	16	32.3	44.3	23.4
Maxillary premolar	12	34.8	39.3	25.9
Mandibular premolar	14	32.1	35.5	32.4
Maxillary central incisor	16	29.2	39.3	31.5

were no exclusionary criteria, and some of the cases treated had significant osseous defects that compromised graft containment. The use of a blended bone allograft to treat these cases provided stability from the osteoconductive capacity of the mineralized cortical component with the additional benefit of improved osteoinductive capacity from the demineralized component.

Notably, while the findings from this case series compare favorably to previously published extraction site preservation studies, some important and significant differences exist. First, the surgical sites in this case series were intentionally left exposed without primary closure. Second, the time allowed for osseous healing from the time of extraction site preservation to the placement of dental implants was shorter than most other studies. These findings suggest that the use of a blended bone allograft product for extraction site preservation procedures has the potential to produce favorable results for the facilitation of future dental implant placement, even in compromised situations.

Conclusion

This case series suggests that excellent results can be obtained when using a blended mineralized-demineralized cortical bone allograft product to preserve alveolar ridge dimensions after tooth extraction. The findings of this case series warrant additional controlled studies with expanded patient populations to confirm these findings.

DISCLOSURE

The author has no affiliation with any of the products mentioned in this article.

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REFERENCES

1. Brown LJ, Babbush CA. The future need and demand for dental implants. In: Babbush CA, Hahn JA, Krauser JT, Rosenlicht JL, eds. *Dental Implants: The Art and Science*. 2nd ed. Maryland Heights, MO: Saunders Elsevier; 2011:1-16.
2. Araújo MG, Lindhe J. Dimensional ridge alterations following tooth extraction. An experimental study in the dog. *J Clin Periodontol*. 2005;32(2):212-218.
3. Amler MH, Johnson PL, Salman I. Histological and histochemical investigation of human alveolar socket healing in undisturbed extraction wounds. *J Am Dent Assoc*. 1960;61:32-44.
4. Pietrokovski J, Massler M. Alveolar ridge resorption following tooth extraction. *J Prosthet Dent*. 1967;17(1):21-27.
5. Hansson S, Halldin A. Alveolar ridge resorption after tooth extraction: A consequence of a fundamental principle of bone physiology. *J Dent Biomech*. 2012;3. doi:10.1177/1758736012456543.
6. Lam RV. Contour changes of the alveolar process following extractions. *J Prosthet Dent*. 1960;10(1):25-32.
7. Schropp L, Wenzel A, Kostopoulos L, Karning T. Bone healing and soft tissue contour changes following single-tooth extraction: a clinical and radiographic 12-month prospective study. *Int J Periodontics Restorative Dent*. 2003;23(4):313-323.
8. Botticelli D, Berglundh T, Lindhe J. Hard-tissue alterations following immediate implant placement in extraction sites. *J Clin Periodontol*.

2004;31(10):820-828.
9. Tarnow DP, Eskow RN, Zamzok J. Aesthetics and implant dentistry. *Periodontol 2000*. 1996;11:85-94.
10. Nevins M, Mellonig JT. Enhancement of the damaged edentulous ridge to receive dental implants: a combination of allograft and the GORE-TEX membrane. *Int J Periodontics Restorative Dent*. 1992;12(2):96-111.
11. Mellonig JR, Triplett RG. Guided tissue regeneration and endosseous dental implants. *Int J Periodontics Restorative Dent*. 1993;13(2):108-119.
12. Simion M, Dahlin C, Trisi P, Piatelli A. Qualitative and quantitative comparative study on different filling materials used in bone tissue regeneration: a controlled clinical study. *Int J Periodontics Restorative Dent*. 1994;14(3):198-215.
13. Lekovic V, Kenney EB, Weinlaender M, et al. A bone regenerative approach to alveolar ridge maintenance following tooth extraction. Report of 10 cases. *J Periodontol*. 1997;68(8):563-570.
14. Iasella JM, Greenwell H, Miller RL, et al. Ridge preservation with freeze-dried bone allograft and a collagen membrane compared to extraction alone for implant site development: a clinical and histologic study in humans. *J Periodontol*. 2003;74(7):990-999.
15. Nevins M, Camelo M, De Paoli S, et al. A study of the fate of the buccal wall of extraction sockets of teeth with prominent roots. *Int J Periodontics Restorative Dent*. 2006;26(1):19-29.
16. Holtzclaw D, Toscano N, Eisenlohr L, Callan D. The safety of bone allografts used in dentistry: a review. *J Am Dent Assoc*. 2008;139(9):1192-1199.
17. Allograft safety and ethical considerations. Proceedings of the fourth symposium sponsored by the Musculoskeletal Transplant Foundation. September 2003. Edinburgh, Scotland, United Kingdom. *Clin Ortho Relat Res*. 2005;435:2-117.
18. Mellonig JT, Bowers GM, Bailey RC. Comparison of bone graft materials. Part I. New Bone formation with autografts and allografts determined by Strontium-85. *J Periodontol*. 1981;52(6):291-296.
19. Mellonig JT, Bowers GM, Cotton WR. Comparison of bone graft materials. Part II. New bone formation with autografts and allografts: a histological evaluation. *J Periodontol*. 1981;52(6):297-302.
20. Toloue SM, Chesnoiu-Matei I, Blanchard SB. A clinical and histomorphometric study of calcium sulfate compared with freeze-dried bone allograft for alveolar ridge preservation. *J Periodontol*. 2012;83(7):847-855.
21. Froum S, Cho SC, Rosenberg E, et al. Histological comparison of healing extraction sockets implanted with bioactive glass or demineralized free-dried bone allograft: a pilot study. *J Periodontol*. 2002;73(1):94-102.
22. Block MS, Finger I, Lytle R. Human mineralized bone in extraction sites before implant placement: preliminary results. *J Am Dent Assoc*. 2002;133(12):1631-1638.
23. Wang HL, Tsao YP. Histologic evaluation of socket augmentation with mineralized human allograft. *Int J Periodontics Restorative Dent*. 2008;28(3):231-237.
24. Fugazzotto PA, Shanaman R, Manos T, Shectman R. Guided bone regeneration around titanium implants: report of the treatment of 1,503 sites with clinical reentries. *Int J Periodontics Restorative Dent*. 1997;17(3):292-299.
25. Committee on Research, Science and Therapy of the American Academy of Periodontology. Tissue Banking of bone allografts used in periodontal regeneration. *J Periodontol*. 2001;72(6):834-838.
26. Urist MR. Bone: formation by autoinduction. *Science*. 1965;150(3698): 893-899.
27. Shigeyama Y, D'Errico JA, Stone R, Somerman MJ. Commercially-prepared allograft material has biological activity in vitro. *J Periodontol*. 1995; 66(6):478-487.
28. Wood RA, Mealey BL. Histologic comparison of healing after tooth extraction with ridge preservation using mineralized versus demineralized free-dried bone allograft. *J Periodontol*. 2012;83(3):329-336.
29. Rosen PS, Reynolds MA. A retrospective case series comparing the use of demineralized freeze-dried bone allograft and freeze-dried bone allograft combined with enamel matrix derivative for the treatment of advanced osseous lesions. *J Periodontol*. 2002;73(8):942-949.
30. Cammack GV II, Nevins M, Clem DS III. Histologic evaluation of mineralized and demineralized freeze-dried bone allograft for ridge and sinus augmentations. *Int J Periodontics Restorative Dent*. 2005;25(3):231-237.

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1. **The routine loss of ridge volume has been explained by the absence of daily stress/strain stimulus required for physiological maintenance of:**
   A. dental implants.
   B. the buccal walls.
   C. the bone anatomy.
   D. bone augmentation.

2. **Various site-preservation techniques have been proposed to reduce the what that occur(s) during post-extraction healing?**
   A. dimensional ridge changes
   B. esthetic changes
   C. pain
   D. amount of swelling

3. **Offering a product that includes bone from the same donor that has been processed in two different ways:**
   A. requires a somewhat more complicated manufacturing process.
   B. can be accomplished in a rather simplified manufacturing process.
   C. has yet to have occurred.
   D. is not allowed by the FDA.

4. **In the case series presented, in all cases primary closure was:**
   A. intentionally achieved.
   B. intentionally not achieved.
   C. unintentionally achieved.
   D. unintentionally not achieved.

5. **After roughly 3 months, bone formation in all cases was adequate to enable proper locational placement of implants without the need for:**
   A. trephine core samples.
   B. an amnion-chorion barrier.
   C. any additional bone-graft material.
   D. a secondary collagen membrane.

6. **The particle size of mineralized bone allograft typically ranges between:**
   A. 25 to 100 micrometers.
   B. 250 to 1,000 micrometers.
   C. 1,250 to 2,500 micrometers.
   D. 2,500 to 10,000 micrometers.

7. **For demineralized bone allograft, the bone is typically immersed in a hydrochloric acid bath in an effort to demineralize the bone by reducing:**
   A. soft tissue.
   B. bacterial contaminants.
   C. bone morphogenetic protein.
   D. calcium content.

8. **The bone allograft used in this case series was a blended ratio of:**
   A. 70:30 demineralized to mineralized cortical bone.
   B. 70:30 mineralized to demineralized cortical bone.
   C. 60:40 mineralized to demineralized cortical bone.
   D. 50:50 demineralized to mineralized cortical bone.

9. **Demineralized bone allograft contains type I collagen and various bone morphogenetic protein (BMP) with the capacity to:**
   A. act as osteoconductive materials.
   B. slow the process of new bone formation.
   C. feature strengthened structural integrity.
   D. influence cell behavior, including proliferation and attachment.

10. **The blended bone allograft provided improved osteoinductive capacity from the demineralized component along with what from the osteoconductive capacity of the mineralized component?**
    A. stability
    B. instability
    C. rapid release of BMPs
    D. compromised graft containment